

Rickettsia felis in *Xenopsylla cheopis*, Java, Indonesia

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Rickettsia typhi and *R. felis*, etiologic agents of murine typhus and fleaborne spotted fever, respectively, were detected in Oriental rat fleas (*Xenopsylla cheopis*) collected from rodents and shrews in Java, Indonesia. We describe the first evidence of *R. felis* in Indonesia and naturally occurring *R. felis* in Oriental rat fleas.

Murine typhus (endemic typhus, fleaborne typhus), caused by *Rickettsia typhi*, is transmitted to humans by infected fleas and is relatively common wherever susceptible rodent hosts reside (1). Fleaborne spotted fever (cat flea typhus), caused by *Rickettsia felis*, is another zoonotic disease carried by fleas and appears to have an equally wide, cosmopolitan distribution; human infections with *R. felis* have a clinical syndrome similar to that of murine typhus (1–4). The cat flea, *Ctenocephalides felis*, has been identified as the primary arthropod vector of *R. felis* in North and South America (United States, Mexico, Peru, Brazil), Europe (Spain, France, United Kingdom, Cyprus), Africa (Gabon, Ethiopia), Asia (Thailand, Afghanistan, Israel), Australia, and New Zealand (1,5–10). We describe the first evidence of *R. felis* in Indonesia and apparent natural infections of *R. felis* in the Oriental rat flea, *Xenopsylla cheopis*, implicating this flea species for the first time as a potential vector for fleaborne spotted fever.

The Study

Samples of *X. cheopis* were collected from 39 live-captured, peridomestic rodents and shrews from 3 localities in Malang, East Java, Indonesia, during an epidemiologic study conducted in 1994 (11). In this study the fleas were reidentified by using morphologic criteria, stored in fresh 70% ethanol, and subsequently evaluated for the presence of rickettsial DNA. DNA sample preparations were derived from triturates of 103 individual fleas in 100 μ L PrepManUltra sample preparation reagent (Applied

Biosystems, Foster City, CA, USA). DNA preparations of 1 to 5 fleas collected from the same rodent were pooled for testing. Reaction mixtures for the quantitative real-time PCR (qPCR) assays had a total volume of 25 μ L and contained 3 μ L DNA template. The master mixes were prepared for the 17-kDa *Rickettsia*-, *R. typhi*- and *R. felis*-specific qPCR assays in a separate, clean (DNA-free) room as previously described (6,12). The primer and probe sequences for the 17-kDa *Rickettsia*-specific and *R. felis*-specific assays have been reported (6,11). The *R. typhi* forward (Rt557F: 5'-TGG TAT TAC TGC TCA ACA AGC T-3') and reverse (Rt678R: 5'-CAG TAA AGT CTA TTG ATC CTA CAC C-3') primers and probe (Rt640BP: 5'-TET-CGC GAT CGT TAA TAG CAG CAC CAG CAT TAT CGC G-DABCYL-3') sequences are listed here. Included in each run were 3 negative controls (GIBCO Ultrapure DNA-free distilled water, Invitrogen Corporation, Grand Island, NY, USA), 1 produced in the clean room and 2 in a biosafety cabinet in another laboratory where DNA templates were added. A TOPO TA plasmid (Invitrogen Corporation) that contained the target sequence at 10³ copies for each assay was used as a positive control. qPCR reactions were incubated in the SmartCycler (Cepheid, Sunnyvale, CA, USA) at 94°C for 2 min, followed by 50 cycles of a 2-step amplification protocol of 94°C for 5 s and 60°C for 30 s. Fluorescence was monitored during the annealing step of each cycle, and data were analyzed with SmartCycler software version 2.0c (Cepheid).

Rickettsial DNA was detected by 17-kDa qPCR in 7 of 39 pools containing 1–5 *X. cheopis* fleas. To determine whether *R. typhi* or *R. felis* infected these fleas, PCR assays specific for *R. typhi* and *R. felis ompB* partial sequence targets were performed (Table). Results of these assays showed that 5 of the 7 *Rickettsia*-positive *X. cheopis* fleas were infected with *R. typhi*, and 2 were positive for *R. felis*. The remainder of the 32 pools and all non-template controls were negative for *R. typhi* and *R. felis*. Additionally, 15 *Rickettsia*-free *C. felis* fleas (Heska Corporation, Loveland, CO, USA), evaluated at the same time and under the same conditions as the Malang fleas, were negative for *R. typhi* and *R. felis*.

Conclusions

To determine the identity of rickettsial agents infecting *X. cheopis* fleas collected from rodents and shrews in Malang, we assessed pools of 1 to 5 fleas from each animal. Our results confirm *R. typhi* in a known flea vector of murine typhus in a highly disease-endemic region of East Java, Indonesia (11,13). *R. felis* has been shown to infect fleas of peridomestic rodents (7,8) and fleas other than *C. felis* (1,5,14). However our report is the first of *R. felis* naturally infecting *X. cheopis* fleas, a vector of plague and

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Table. Detection of *Rickettsia typhi* and *Rickettsia felis* in flea pools by quantitative, real-time PCR, East Java, Indonesia

Location and distribution of rodents and shrews	No. flea pools (total no. fleas)*	No. positive pools (%)		
		17 kDa	<i>R. typhi</i>	<i>R. felis</i>
Rural (Mulyorejo)				
<i>Rattus argentiventer</i> (7.1%)	1 (2)	0	0	0
<i>Rattus rattus</i> (14.3%)	2 (3)	0	0	0
<i>Rattus tiomanicus</i> (57.1%)	1 (1)	0	0	0
<i>Mus musculus</i> (10.7%)	1 (2)	0	0	0
<i>Chiropodomys gliroides</i> (3.6%)	0			
<i>Suncus murinus</i> (7.1%)	0			
Total rural	5 (8)	0	0	0
Suburban (Bandungrejosari)				
<i>Rattus exulans</i> (3.2%)	1 (3)	0	0	0
<i>R. rattus</i> (74.2%)	13 (32)	1	0	1
<i>R. tiomanicus</i> (19.4%)	4 (9)	0	0	0
<i>C. gliroides</i> (3.2%)	1 (4)	0	0	0
Total suburban	19 (48)	1 (5.3)	0	1 (5.3)
Urban (Klojen)				
<i>R. exulans</i> (5.3%)	1 (4)	1	1	0
<i>R. rattus</i> (68.4%)	9 (28)	4	3	1
<i>R. tiomanicus</i> (10.5%)	2 (7)	0	0	0
<i>Rattus sabanus</i> (10.5%)	2 (5)	0	0	0
<i>M. musculus</i> (5.3%)	1 (3)	1	1	0
Total urban	15 (47)	6 (40)	5 (33.3)	1 (6.7)
Total	39 (103)	7 (18)	5 (12.8)	2 (5.1)

*Each pool of 1 to 5 fleas (*Xenopsylla cheopis*) came from a single animal.

murine typhus. Both *R. felis*-containing flea pools were derived from *Rattus rattus*, 1 from the suburban and 1 from the urban neighborhoods of Malang. *R. rattus* was the predominant species captured in urban and suburban environments (72%) and appears to be the primary host for *R. felis*- and *R. typhi*-infected *X. cheopis*. In the rural setting, where *R. rattus* was represented with far less frequency (14.3%), neither rickettsial agent was detected in collected fleas. These findings merit further epidemiologic investigation to better understand the relationship between *R. felis*, *R. typhi*, and *X. cheopis* and the transmission dynamics between flea and rodent.

Additionally, this report provides the first evidence of *R. felis* in the Indonesian archipelago. Investigations of rickettsial agents in Indonesia have been relatively few; to date, human infections with *R. felis* have not been reported from Indonesia. The lack of reports may be because a murine typhus-like disease associated with *R. felis* infection would not allow healthcare providers to clinically discriminate fleaborne spotted fever from murine typhus or other rickettsioses. In Indonesia, rickettsioses and typhoid fever are collectively referred to as *tifus*. Rickettsial tifus can be discerned by serologic tests or by observing when rickettsial tifus cases rapidly respond to treatment with a tetracycline or chloramphenicol. Furthermore, the inability to diagnose fleaborne spotted fever by laboratory means has been attributed to the cross-reactivity of antibodies to *R. felis* antigens with other rickettsial antigens (1). Consequently, serologic assays have been unable to differ-

entiate fleaborne spotted fever from other rickettsioses. Thus, the high prevalence of murine typhus reported in Indonesia likely also includes fleaborne spotted fever. In addition, previously demonstrated serologic evidence of spotted fever group rickettsiae infection among residents of Gag Island, in eastern Indonesia (15), could have been due to *R. felis*. On the basis of data presented here and of recent reports of *R. felis* in other countries in Asia (2–5,8,9), healthcare providers in Indonesia should be alerted to the possibility of fleaborne spotted fever among their patients.

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Dr Jiang works at the Rickettsial Diseases Department of the Naval Medical Research Center, Silver Spring, Maryland. Her research interests include rickettsial epidemiology, host immune response to rickettsial infection, and rapid diagnostic assay and vaccine development.

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